

## FLUCTUATION OF POST-PRANDIAL PLASMA MINERAL LEVEL OF JUVENILE JAPANESE FLOUNDER, *Paralichthys olivaceus* FED DIETARY PHOSPHORUS AND PHYTASE SUPPLEMENTATION

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### ABSTRACT

In order to investigate the phytic acid degradation in the gut of post juvenile Japanese flounder, indirect method was carried out by measuring the pre-prandial and post-prandial plasma mineral and alkaline phosphatase (ALP) level as well as liver phosphorus content. The experiment was designed into a Randomized Block in which experiment units were grouped according to sampling days at 10, 20 and 30 days of feeding time. Experimental diets contained three levels of dietary inorganic phosphorus at 0.0; 0.25 and 0.5% combined with two levels of dietary phytase at 0 and 2000 FTU/kg diet. Juvenile Japanese flounder (IBW = 36.2 g) were randomly distributed into 6 tanks of a 200 L capacity with density of 15 fish/tank. Blood sampling was carried out at 0 hour (before feeding or pre-prandial) and at 1, 3, 6 and 12 hour post feeding (post-prandial) time in three sampling days, respectively. Plasma was measured for mineral and ALP levels, while liver was analyzed for P content. The observation showed that fish fed without both dietary IP and phytase supplements had the lowest postprandial plasma IP, Mg and ALP levels during 12-h postprandial period. Plasma IP level at 6-h post-feeding in groups fed dietary 0.25 and 0.5% IP were significant higher when diet supplemented with phytase than those without phytase supplement. Peak level of plasma IP in fish fed 0.25% IP was similar to fish fed 0.5% with the presence of dietary phytase. At 1 and 3-h post-feeding, plasma Ca level increased in all groups, but significant difference was only observed between group fed diet without both dietary IP and phytase and other groups. Similar to plasma IP level, peak of plasma Mg and ALP concentration occurred in fish fed 0.25% IP together with phytase, and did not significantly differ from fish fed with 0.5% IP even when phytase was included in diet.

**KEYWORDS:** phytase, phosphorus, mineral, alkaline phosphatase, Japanese flounder

### INTRODUCTION

Phytase presents widely in plants, certain animal tissues and microorganism particularly fungi. Common plants used as ingredient for

fish nutrition such as soybean, canola, maize and wheat contain phytase, but has limited importance due to low activity in the feed ingredients and inactivation by heat treatment during processing (Phillippy, 2001). Thus, de-

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phosphorylation of phytic acid or also called myo-inositol hexakiphosphates (IP6) in the digestive tract of monogastric animals such as chicken, pig and fish is believed mainly due to the supplemented phytase in diet. In addition, the non-enzymatic degradation of IP6 in the stomach can be considered as negligible, because IP6 molecule is very stable when exposed to gastric pH condition (Kemme *et al.*, 1998).

Phytase commonly used in animal diet including fish diet is categorized acid phytase with pH spectrum varying according to phytase sources. The commercial phytase isolated from *Peniophora lycii* used in this study has optimal pH activity at pH 4.0-5.0 (Pontoppidan *et al.*, 2007) which is in accordance with the pH of *in vitro* IP6 degradation occurred at pH 5.0 and 4.0 with less degradation occurring at pH 3.0. The pH of the gastrointestinal tract has been reported as one of the most critical parameters for *in vivo* degradation of IP6. Degradation of phosphorus (P) associated IP6 (phytate-P) is generally thought to proceed during crop digestion in chicken where pH is in a range of 4.0 to 6.5 (Sarra *et al.*, 1992; Selle & Ravindran, 2007) and during gastric digestion in pig where the pH ranges from 2.0-5.0 (Argenzio & Southworth, 1975; Kemme *et al.*, 2006). In case of fish, it is most likely that degradation of IP6 occurs in stomach since the pH of stomach is between 2.0 and 4.0 depending on the species. Sugiura & Ferraris (2004) reported that stomach of rainbow trout filled with feed had pH of chime around 4 and remained at the same unit even 6-9 h after feeding. Similarly, stomach pH of juvenile Japanese flounder after 1 h feeding was around 4.19 and became alkaline in intestine (Laining *et al.*, 2010a).

Degradation and formation of IP6 during digestion is a subject that has scarcely been touched particularly in case of fish. The direct method to determine the IP6 degradation in fish gut is to collect the digestive contents and then analyze for phytase activity, IP6 and its degradation products. However, one of the difficulties in analyzing these parameters is the limit amount of digestive contents, particularly in small fish to meet the samples need. Alternatively, indirect method to determine the IP6 degradation in fish is to measure the postprandial plasma inorganic P (IP) concentration and other minerals (Liebert & Portz, 2007).

Mineral content of diet has been reported to be one of the most critical factors affecting

the efficacy of phytase *in vivo* in poultry Angel *et al.* (2002) and swine (Seynaeve *et al.*, 2000). However, limit information is available in fish species including our previous study dealing with juvenile tiger puffer (Laining *et al.*, 2010b). In regard to growth performances, dietary Ca/P ratio of 0.5 or without additional Ca supplement combined with 2000 FTU phytase per kg might be the optimum combination in diet for tiger puffer. Furthermore, dietary phytase and IP levels had shown an independent effect on growth in juvenile red sea bream and interaction effect was found on P digestibility, vertebral Zn and scale P content (Laining *et al.*, 2010c). Based on identical diet used for growth trial in juvenile Japanese flounder, this present study aimed at investigating the degradation of IP6 in the gut of post juvenile Japanese flounder along with corresponding postprandial plasma mineral and alkaline phosphatase (ALP) concentration.

## MATERIALS AND METHODS

### Experimental Design

This experiment was designed into a Randomized Block Design in which experimental units were grouped (blocked) according to the sampling time and the treatments were assigned to the experimental units within the block by random. Sampling was carried out three times in every 10 days for 30 days. Formulation of experimental diet was similar to diet used in the previous study (Table 1). Based on analysis, composition of the experimental diet is presented in Table 2).

### Observation on Change of Postprandial Plasma Mineral and ALP level

Post juvenile Japanese flounder were obtained from a commercial hatchery (Matsumoto Suisan Co., Miyazaki, Japan) and maintained with commercial diet (Higashimaru, Kagoshima, Japan) for one week. Fish with average IBW of 36.2 g were distributed into a 6 tanks of 200 L capacity at a density of 15 fish/tank. Each tank was equipped with continuous aeration and supplied with seawater with flow through system (2 L/minutes). Fish were fed the experimental diet twice a day to nearly satiation in the morning at 08 00 and in the afternoon at 16 00. After ten days feeding, in following day blood sampling was carried out at 0-h (before feeding/ pre-prandial) and at 1, 3, 6 and 12-h

Table 1. Formulation of experimental diet contained different level of dietary IP and microbial phytase

Ingredients (%)	Phosphorus (P) / Phytase level (P = phytase; NP = non phytase)					
	0/NP	0/P	0.25/NP	0.25/P	0.5/NP	0.5/P
Brown fishmeal	18	18	18	18	18	18
Krill meal	10	10	10	10	10	10
Soybean protein isolated <sup>1</sup>	30	30	30	30	30	30
Dextrin-hydrate	10	10	10	10	5	5
$\alpha$ -Starch	5	5	5	5	5	5
Pollack liver oil	8	8	8	8	8	8
HUFA <sup>2</sup>	1	1	1	1	1	1
Activated gluten	5	5	5	5	5	5
Vitamin mix <sup>3</sup>	4	4	4	4	4	4
Stay C <sup>4</sup>	0.1	0.1	0.1	0.1	0.1	0.1
Ca-P-free mineral mix <sup>5</sup>	4	4	4	4	4	4
Sodium monophosphate <sup>6</sup>	0	0	1.1	1.1	2.2	2.2
Calcium carbonate	0.5	0.5	0.5	0.5	0.5	0.5
Phytase <sup>7</sup>	0	0.04	0	0.04	0	0.04
Betaine <sup>8</sup>	0.2	0.2	0.2	0.2	0.2	0.2
$\alpha$ -Cellulose	4.2	4.16	3.1	3.06	2	1.96

<sup>1</sup> Soybean protein isolated obtained from Fuji Pro Company, Tokyo, Japan

<sup>2</sup> Poweash A, Oriental Yeast Co, Ltd, Tokyo, Japan

<sup>3</sup> Vitamin mixture (g kg<sup>-1</sup> diet):  $\beta$ -carotene 0.192; vitamin D3 0.019; menadione 0.0917;  $\alpha$ -tocopherol acetate 0.77; thiamin nitrate 0.115; riboflavin 0.385; pyridoxine-HCl 0.092; cyanocobalamin 0.00018; d-biotin 0.0115; inositol 7.698; nicotinic acid 1.539; Ca-pantothenate 0.5391; folic acid 0.0288; choline chloride 15.738; p-aminobenzoic acid 0.7665; cellulose 2.849

<sup>4</sup> L-ascorbyl-Na/Ca (DSM Nutritional Product Ltd, Basel, Switzerland)

<sup>5</sup> Calcium-phosphorus free mineral mixture (g/kg diet): KCl 1.856; MgSO<sub>4</sub>·5H<sub>2</sub>O 5.067; Fe Citrate 1.098; Al (OH)<sub>3</sub> 0.0069; ZnSO<sub>4</sub>·7H<sub>2</sub>O 0.132; CuSO<sub>4</sub> 0.0037; MnSO<sub>4</sub>·5H<sub>2</sub>O 0.029; K (IO<sub>3</sub>)<sub>2</sub> 0.006; CoSO<sub>4</sub>·7H<sub>2</sub>O 0.037; Cellulose 31.75

<sup>6</sup> Wako Pure Chemical Industries, Ltd, Japan

<sup>7</sup> Ronozyme P5000, DSM Nutritional Product Ltd, Basel, Switzerland (declared activity = 5000 FTU g<sup>-1</sup> product)

<sup>8</sup> Attractant (g kg<sup>-1</sup> diet): betaine 2.0

post feeding time (post-prandial). One fish from each tank was taken for blood sampling and sacrificed for liver sample. Following sampling at 20 days and 30 days of feeding time were done accordingly. Blood was immediately centrifuged (4000 rpm, 10 min, 4°C) to separate the plasma. All samples were stored in -80°C until analysis.

### Chemical Analysis

Analysis of IP, Ca, Mg and ALP in plasma was carried out by using a blood analyzer

(Spotchem™ EZ SP 4430, Arkray Inc., Kyoto, Japan). Total P in the liver was analyzed according to Lowry & Lopez (1946).

### Statistical Analysis

Data were statistically analyzed by ANOVA for Randomized Block using SPSS software package (Version 15 for Windows; SPSS Inc., Chicago, IL, USA). Duncan multiple range post-hoc test was performed to identify the differences among treatments. The statistical significance was set at P < 0.05.

Table 2. Chemical composition of experimental diet (%)

Chemical composition	Inorganic P (IP) / Phytase level (P = phytase; NP= non phytase)					
	OIP/NP	OIP/P	0.25IP/NP	0.25IP/P	0.5IP/NP	0.5/P
Moisture	7.7	8	7.4	8	7.3	7.3
Crude protein	48.9	49.2	48	48	48.5	49
Total lipid	12	12.6	12.5	12.3	12	12.3
Ash	7.1	7.2	7.9	7.9	8.6	8.6
Phytic acid/IP6	0.53	0.5	0.54	0.52	0.55	0.56
Minerals content:						
- Total P (%)	0.82	0.84	1.03	1.05	1.24	1.17
- Ca (%)	0.87	0.82	0.78	0.87	0.84	0.83
- Mg (mg/g)	1.71	1.66	1.4	1.54	1.6	1.5
- Zn (mg/g)	69.14	69.01	61.4	67.25	69.57	65.87
- Ca/P ratio	0.94	0.98	0.8	0.83	0.68	0.71

**RESULTS AND DISCUSSION**

**Result**

***Change of postprandial plasma IP, Ca and Mg***

Plasma IP concentration during pre-prandial period and during 12-h postprandial period at three sampling days is presented in Table 3. Mean pre-prandial plasma IP level was significantly higher in two groups fed the optimum level of dietary IP (with and without phytase supplement) compared to other four groups fed lower levels of dietary IP (Figure 1). After feeding, concentrations increased within 1-h and reached the peak concentration at 3 and 6-h depending on the treatments. Fish fed without both dietary IP and supplemental phytase had the lowest postprandial plasma IP level among groups in all sampling times. Sampling at 6-h post feeding, plasma IP concentration in groups fed dietary IP (0.25% and 0.5%) were significant higher when diet supplemented with phytase than those without phytase supplementation. When dietary phytase was included in the diet, peak concentration was achieved in fish fed 0.25% IP (22.2 mg/dL), but did not significantly differ from fish fed 0.5% IP (21.0 mg/dL). The 12-h post-prandial IP levels showed the decreasing peak of P absorption.

Pre and postprandial plasma Ca levels are shown in Table 4. There was no clear trend on the mean pre-prandial plasma Ca concentra-

tion among treatments as illustrated in Figure 2. Sampling at 1 and 3-h post-feeding showed an increasing Ca level in all groups but significant difference was only observed between group fed diet contained no dietary IP and phytase and another 5 groups. The peak Ca concentration occurred at 1 and 3-h after feeding depending on treatments. Decreasing peak of Ca level was shown at 6 and 12-h post-feeding.

Table 5 presents plasma Mg level during pre-prandial and 12-h post-prandial period. Similar to plasma IP, pre-prandial Mg was significantly higher in groups fed diet with 0.5% IP at two phytase levels and in group fed 0.25% IP with phytase compared to other groups fed lower dietary IP. The peak concentration was achieved after 3-h post-feeding except in fish fed diet without both dietary IP and phytase which its peak occurred at 6-h after feeding (Figure 3). The peak Mg concentration was highest in fish fed 0.25% IP together with phytase but did not significantly differ from two groups fed 0.5% IP.

Sampling at 10, 20, and 30 days of feeding time (as the block) did not significantly affect the pre-prandial and 12-h postprandial plasma IP, Ca and Mg.

***Change of post-prandial plasma ALP***

Concentration of ALP during pre-prandial and 12-h post-prandial period is shown in Table

Table 3. Change of post-prandial plasma IP level of Japanese flounder fed different levels of dietary IP and phytase supplementation over 12-h at three samplings of each treatment

Treatment	Day	Postprandial IP (mg/dL)												
		0-h	Mean	1-h	Mean	3-h	Mean	6-h	Mean	12-h	Mean			
0IP/NP	10-d	6.1		15.4		15.6		17.6		13.5		16.0		11.6 <sup>ab</sup>
	20-d	9.2	7.1 <sup>#</sup>	13.1	13.1 <sup>a</sup>	15.6	14.5 <sup>a</sup>	16.0	16.5 <sup>a</sup>	10.4	16.0	16.0	11.0	
	30-d	6.1		10.8		12.4		18.0		13.4		20.0		14.1 <sup>bc</sup>
0IP/P	10-d	6.6		15.1		19.8		18.0		14.8		18.0		14.0
	20-d	10.8	8.1 <sup>a</sup>	17.6	17.6 <sup>b</sup>	19.9	19.9 <sup>b</sup>	20.0	18.7 <sup>ab</sup>	14.8	20.0	18.0	14.0	
	30-d	6.8		20.0		20.0		17.2		12.6		19.1		12.6 <sup>abc</sup>
0.25IP/NP	10-d	9.8		13.0		18.2		17.2		10.9		18.8		14.4
	20-d	7.7	8.8 <sup>a</sup>	12.9	13.0 <sup>a</sup>	19.4	19.2 <sup>b</sup>	20.0	18.4 <sup>ab</sup>	10.9	19.1	18.8	14.4	
	30-d	8.9		13.0		20.0		22.2		15.3		20.0		17.2 <sup>d</sup>
0.25IP/P	10-d	10.9		17.7		20.0		20.0		17.9		24.4		18.4
	20-d	13.3	12.1 <sup>b</sup>	20.0	19.2 <sup>b</sup>	20.0	20.0 <sup>b</sup>	20.0	22.2 <sup>c</sup>	10.2	17.7	16.6	10.3	
	30-d	12.2		20.0		20.0		19.7 <sup>b</sup>		17.7		21.0		15.2 <sup>cd</sup>
0.5IP/NP	10-d	14.4		15.7		19.1		18.9		15.0		20.1		13.4
	20-d	12.6	13.8 <sup>bc</sup>	20.0	18.6 <sup>b</sup>	20.0	19.1 <sup>b</sup>	20.1	17.7 <sup>a</sup>	10.0	17.7	16.6	10.3	
	30-d	14.4		20.0		20.0		19.1		17.2		22.0		13.4
0.5IP/P	10-d	16.7		17.8		19.1		20.1		15.0		21.0		13.4
	20-d	15.3	16.0 <sup>c</sup>	20.0	18.9 <sup>b</sup>	19.2	19.1 <sup>b</sup>	21.0	21.0 <sup>bc</sup>	17.2	21.0	22.0	13.4	
	30-d	16.0		18.9		19.0		19.0		17.2		17.2		13.4

**ANOVA of Randomized Block**

Source	Postprandial IP (mg/dL)					
	0-h	1-h	3-h	6-h	12-h	
Treatment Block (day of sampling)	*	*	*	*	*	*
	NS	NS	NS	NS	NS	NS

# Values in the each column with same superscripts are not significantly different (P>0.05)

\* : Significant P<0.05; NS: no significant

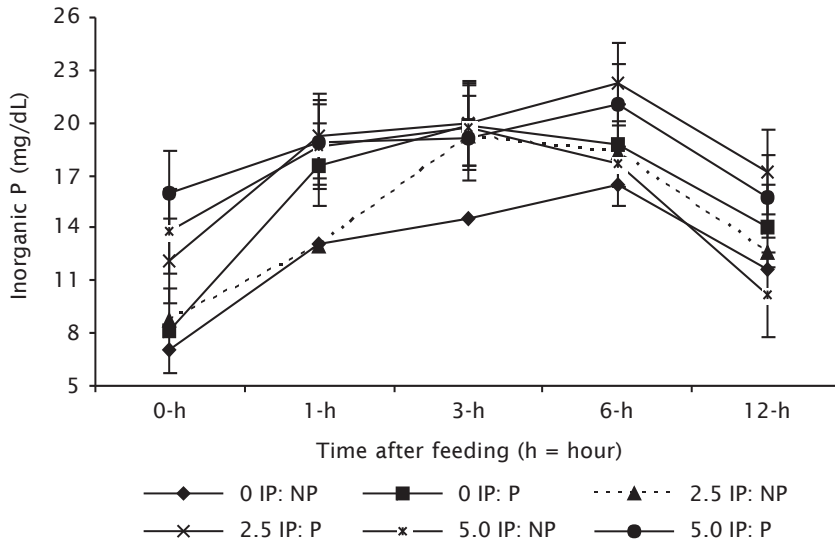


Figure 1. Change of postprandial plasma IP of Japanese flounder fed different levels of dietary IP and phytase supplementation over 12-h. Each point represents the mean values ( $\pm$  SE) of three samplings of each treatment

6. Pre-prandial plasma ALP was significantly higher in two groups fed 0.5% IP than other four groups fed lower level of IP. Generally, fish fed without both dietary IP and phytase supplementation had significantly lower ALP level during 12-h postprandial period as indicated in Figure 4. Increasing of plasma ALP concentration occurred at 1-h postfeeding except in group fed diet without both IP and phytase supplement. Concentration of ALP achieved the peak at 6-h post-feeding in all groups except in group fed diet with 0.25% IP without supplemental phytase. The highest peak of ALP level was in group fed diet with 0.25% IP and phytase, but did not significantly differ from three other groups (group fed no IP with phytase and two groups fed higher dietary IP of 0.5%).

#### Liver P content at 0, 6, and 12-h postprandial

Change of P content in liver at pre and 12-h post-prandial period is presented in Table 7. Pre-prandial liver P content was not significant different in four groups fed diet without dietary IP and 0.25% IP at two levels of dietary phytase. However, there was significant different between fish fed no dietary IP and 0.5% IP. Similar trend was observed at 6-h post-feeding. At 12-

h post-feeding, concentration of P in liver was significant higher in fish 0.5% IP, but did not improve with the presence of phytase in diet.

Based on statistical analysis, there was a significant effect of sampling days on P liver content in which sampling at 10 days was significant different from sampling at 20 and 30 days.

#### Discussion

Incorporation of dietary phytase in fish diets have been reported to improve P availability and growth performances (Rodehutsord & Pfeffer, 1995; Yan & Reigh, 2002). However, the observation of the specific effects of phytase in different fish species is insufficient. The result of this present study demonstrated the different effect of dietary IP and phytase levels on concentration of mineral and ALP in plasma as well as liver P content which are further explained by physiological data on the change of 12-h post-prandial of these parameters. Lower plasma IP in fish fed diet without both dietary IP and phytase at pre and 12-h post-prandial period indicated that absorption of IP from this diet in digestive tract of red sea bream was very low compared to other groups. This result revealed that P originated from or-

Table 4. Change of post-prandial Ca of Japanese flounder fed different levels of dietary IP and phytase supplementation over 12-h at three samplings of each treatment

Treatment	Day	Postprandial Ca (mg/dL)													
		0-h			1-h			3-h			6-h			12-h	
		Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
0IP/NP	10-d	11.2		12.8		13.2		12.0		13.3		12.0		13.3	
	20-d	13.6		14.5		16.4		15.2 <sup>a</sup>		12.6 <sup>a</sup>		13.8		14.4	
	30-d	14.2		11.1		14.0		17.0 <sup>a</sup>		16.9 <sup>b</sup>		11.9		12.1	
0IP/P	10-d	10.2		19		16.2		15.4		11.4		15.4		11.4	
	20-d	12.8		18.4		16.2		18.8 <sup>c</sup>		14.3		16.9		14.3	
	30-d	11.3		18.9		18.7		17.0 <sup>a</sup>		16.9 <sup>b</sup>		18.4		14.4	
0.25IP/NP	10-d	9.70		16.3		14.9		14.9		14.7		14.9		14.7	
	20-d	11.2		17.8		16.6		16.4 <sup>a</sup>		15.9 <sup>b</sup>		17.2		14.5	
	30-d	11.0		14.8		17.7		15.5		14.6		15.5		14.6	
0.25IP/P	10-d	15.7		17		17.9		14.5		14.8		14.5		14.8	
	20-d	16.6		15.7		18.7		17.9 <sup>a</sup>		15.6 <sup>b</sup>		15.8		12.4	
	30-d	14.7		18.7		17.1		16.6		13.6		16.6		13.6	
0.5IP/NP	10-d	15.7		16.2		17.6		12.7		10.8		12.7		10.8	
	20-d	16.4		15.8		18.8		17.6 <sup>a</sup>		13.4 <sup>a</sup>		14.2		11.5	
	30-d	15.0		16.6		16.5		13.4		10.9		13.4		10.9	
0.5IP/P	10-d	10.3		14.1		17.5		13.5		10.6		13.5		10.6	
	20-d	12.5		17.5		14.7		16.2 <sup>a</sup>		13.5 <sup>a</sup>		12.7		10.8	
	30-d	14.6		16.8		16.3		14.3		11.0		14.3		11.0	

**ANOVA of Randomized Block**

Source	Postprandial Ca (mg/dL)					
	0-h	1-h	3-h	6-h	12-h	
Treatments Block (day of sampling)	*	*	*	*	*	*
	NS	NS	NS	NS	NS	NS

# Values in the each column with same superscripts are not significantly different (P>0.05)

\* : Significant P< 0.05; NS: no significant

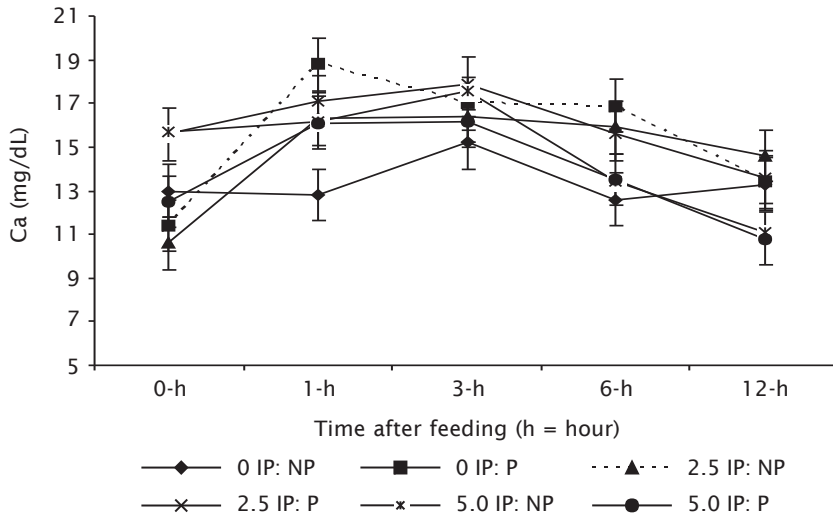


Figure 2 . Change of postprandial plasma Ca of Japanese flounder fed different levels of dietary IP and phytase supplementation over 12-h. Each poin represents the mean values ( $\pm$  SE) of three samplings of each treatment

ganic sources of fishmeal, krill meal and SPI in diet is not effectively digested by fish. The main form of dietary P in fish diet is hydroxyapatite or bone phosphate which requires strong acidity to be solubilized in the stomach for subsequent absorption in the intestine (Sugiura *et al.*, 2006). This may be the reason why P in fishmeal is well-digested by mammals (Soares Jr, 1995) which have very low post-prandial stomach pH of 2 or less (Benne & Levy, 2000) and poorly digested by fish (Cho & Bureau, 2001). Moreover, fish cannot effectively utilize the P associated to IP6 coming from SPI since phytase was not included in this group. Significant higher plasma IP level in fish fed dietary IP which was clearly shown at 3-h post-feeding time revealed that availability of P from inorganic source is more available than organic source. Resent study dealing with yellow tail reported that sodium phosphate monobasic as used in this experiment had much higher digestibility compared to tribasic form which is similar to phosphate form in fishmeal (Sarker *et al.*, 2009). Furthermore, presence of phytase in diet significantly elevated the plasma IP level particularly at 6-h post-feeding time indicating the higher P availability originated from IP6 after being degraded by phytase. At 6-h blood sampling, supplementation of phytase in diet contained 0.25% IP had significant higher peak

value than fish fed no dietary IP, but did not further improve when dietary IP increased to 0.5%.

Generally, P metabolism in fish is thought to be similar to that in mammals. Sugiura *et al.* (2003) reported that IP is transported by IP transporter in the apical membrane of intestinal cells, as well as paracellular transporter-independent pathways, and subsequently transported into plasma. Thus, IP that is not absorbed by intestine is secreted in feces. If it is not reabsorbed by IP transporter in the renal proximal tubule, plasma IP diffuses into urine in kidney and is excreted with the urine. In the present study, even though intestinal P absorption was not directly measured, higher plasma IP level in fish fed diet with phytase supplement provided evidence for a faster P uptake in the digestive tract of fish compared to unsupplemented groups. To our knowledge, only one study is so far available reporting the post-feeding time course of IP in blood plasma of Nile tilapia (Liebert & Portz, 2007). The authors found that following the highest dietary phytase at 1000 and 1250 FTU/kg diet, blood sampling at 2-h showed a similar response on plasma IP as the control diet contained IP supplement and become more pronounced at 4-h post-feeding time. In addition, they observed that the peak of plasma IP level oc-



Table 5. Change of postprandial Mg of Japanese flounder fed different levels of dietary IP and phytase supplementation over 12-h at three samplings of each treatment

Treatment	Day	Postprandial Mg (mg/dL)												
		0-h	Mean	1-h	Mean	3-h	Mean	6-h	Mean	12-h	Mean	12-h	Mean	
0IP/NP	10-d	2.2		2.5		2.0		2.3		2.0		2.3		1.6
	20-d	2.3	2.3 <sup>ab</sup>	2.7	2.5 <sup>a</sup>	2.2	2.3 <sup>a</sup>	2.9	2.9 <sup>b</sup>	2.2	2.3 <sup>a</sup>	2.9	2.9 <sup>b</sup>	2.6
	30-d	2.3		2.3		2.6		3.5		2.6		3.5		2.0
0IP/P	10-d	2.7		3.7		3.9		3.1		3.9		3.1		2.9
	20-d	2.6	2.6 <sup>ab</sup>	2.5	3.3 <sup>bc</sup>	3.0	3.5 <sup>b</sup>	3.0	3.1 <sup>bc</sup>	3.0	3.5 <sup>b</sup>	3.0	3.1 <sup>bc</sup>	2.4
	30-d	2.4		3.7		3.5		3.1		3.5		3.1		3.5
0.25IP/NP	10-d	2.4		2.5		4.2		2.6		4.2		2.6		2.2
	20-d	2.6	2.4 <sup>a</sup>	2.9	2.7 <sup>ab</sup>	4.4	4.5 <sup>c</sup>	3.2	3.2 <sup>bc</sup>	4.4	4.5 <sup>c</sup>	3.2	3.2 <sup>bc</sup>	2.6
	30-d	2.1		2.8		4.8		3.5		4.8		3.5		2.4
0.25IP/P	10-d	3.0		3.7		4.7		3.6		4.7		3.6		2.7
	20-d	3.3	3.0 <sup>b</sup>	3.8	3.8 <sup>c</sup>	5.6	5.2 <sup>d</sup>	3.2	3.5 <sup>c</sup>	5.6	5.2 <sup>d</sup>	3.2	3.5 <sup>c</sup>	2.9
	30-d	2.6		4.0		5.4		3.7		5.4		3.7		3.4
0.5IP/NP	10-d	3.1		3.1		4.0		2.4		4.0		2.4		2.4
	20-d	2.7	3.1 <sup>b</sup>	2.5	3.1 <sup>abc</sup>	4.6	4.5 <sup>cd</sup>	2.4	2.4 <sup>a</sup>	4.6	4.5 <sup>cd</sup>	2.4	2.4 <sup>a</sup>	2.5
	30-d	3.5		3.6		5.0		2.3		5.0		2.3		2.7
0.5IP/P	10-d	2.9		2.9		4.2		3.0		4.2		3.0		2.1
	20-d	2.7	2.9 <sup>b</sup>	3.4	3.2 <sup>abc</sup>	4.9	4.8 <sup>cd</sup>	3.7	3.5 <sup>c</sup>	4.9	4.8 <sup>cd</sup>	3.7	3.5 <sup>c</sup>	2.7
	30-d	3.1		3.2		5.4		3.7		5.4		3.7		2.4

**ANOVA of Randomized Block**

Source	Postprandial Ca (mg/dL)					
	0-h	1-h	3-h	6-h	12-h	12-h
Treatments	*	*	*	*	*	*
Block (day of sampling)	NS	NS	NS	NS	NS	NS

# Values in the each column with same superscripts are not significantly different (P>0.05)

\* : Significant P < 0.05; NS: no significant

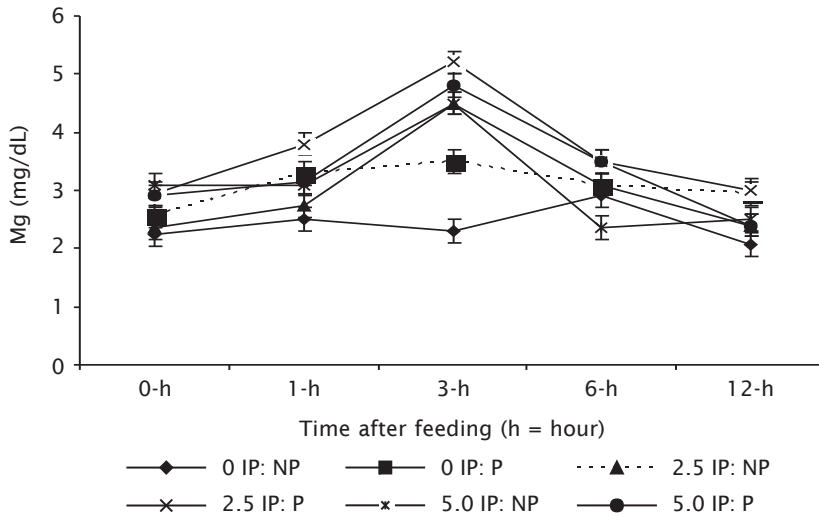


Figure 3. Change of postprandial plasma Mg of Japanese flounder fed different levels of dietary IP and phytase supplementation over 12-h. Each point represents the mean values ( $\pm$  SE) of three samplings of each treatment

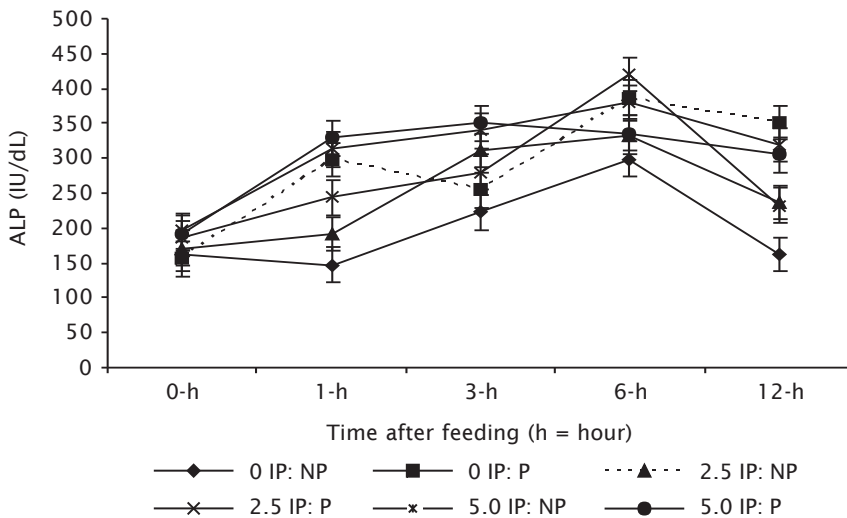


Figure 4. Change of postprandial plasma ALP of Japanese flounder fed different levels of dietary IP and phytase supplementation over 12-h. Each point represents the mean values ( $\pm$  SEM) of three samplings of each treatment

curred at 8-h post-feeding and tended to exceed the effect of feeding control IP diet. Nile tilapia seemed to require lower phytase level to achieve the peak of plasma IP level compared to Japanese flounder as indicated in this study.

Based on statistical analysis, sampling days which reflected the fish age did not significantly affect the plasma IP level. Satoh *et al.* (2002) reported that in rainbow trout, P absorption from fishmeal decreased slightly as body weight increased. This was in contrast to P

Table 6. Change of postprandial ALP of Japanese flounder fed different levels of dietary IP and phytase supplementation over 12-h at three samplings of each treatment

Treatment	Day	Postprandial ALP (IU/dL)												
		0-h	Mean	1-h	Mean	3-h	Mean	6-h	Mean	12-h	Mean			
0IP/NP	10-d	130		130		177		154		130		154		130
	20-d	177	163.0 <sup>#</sup>	132	146.7 <sup>a</sup>	270	223.0 <sup>a</sup>	338	263.6 <sup>a</sup>	181	162.0 <sup>a</sup>	338	263.6 <sup>a</sup>	181
	30-d	182		178		222		298.7		175		298.7		175
0IP/P	10-d	150		256		223		338		292		338		292
	20-d	144	156.0 <sup>a</sup>	255	298 <sup>cd</sup>	255	255.0 <sup>ab</sup>	420	383.3 <sup>b</sup>	332	351.0 <sup>c</sup>	420	383.3 <sup>b</sup>	332
	30-d	175		383		287		400		351		400		351
0.25IP/NP	10-d	130		192		304		202		130		202		130
	20-d	206	171.3 <sup>a</sup>	192	207.2 <sup>ab</sup>	282	311.3 <sup>cd</sup>	301	289.7 <sup>ab</sup>	271	236.7 <sup>b</sup>	301	289.7 <sup>ab</sup>	271
	30-d	178		238		348		332		309		332		309
0.25IP/P	10-d	199		230		253		420		232		420		232
	20-d	172	185.7 <sup>ab</sup>	258	244 <sup>bc</sup>	312	280.3 <sup>bc</sup>	532	459.7 <sup>c</sup>	242	232.0 <sup>b</sup>	532	459.7 <sup>c</sup>	242
	30-d	186		244		276		427		222		427		222
0.5IP/NP	10-d	228		267		288		347		274		347		274
	20-d	218	228.5 <sup>c</sup>	267	313.3 <sup>cd</sup>	373	339.3 <sup>d</sup>	362	379.7 <sup>bc</sup>	305	319.3 <sup>c</sup>	362	379.7 <sup>bc</sup>	305
	30-d	239		406		357		430		379		430		379
0.5IP/P	10-d	198		288		295		367		252		367		252
	20-d	224	224.0 <sup>bc</sup>	251	329 <sup>d</sup>	350	350.3 <sup>a</sup>	377.7	386.9 <sup>bc</sup>	259	305.3 <sup>c</sup>	377.7	386.9 <sup>bc</sup>	259
	30-d	250		448		406		293		305		293		305

**ANOVA of Randomized Block**

Source	Postprandial Ca (mg/dL)			
	0-h	1-h	3-h	6-h
Block (day of sampling)	NS	*	*	*
10 days		a	a	a
20 days		a	b	b
30 days		b	b	b

# Values in the each column with same superscripts are not significantly different (P>0.05)

\* : Significant P < 0.05; NS: no significant

Table 7. Liver total P content of Japanese flounder before (0-h) and after 6-h and 12-h feeding diet containing different levels of dietary IP and phytase supplementation at three sampling days

Treatment	Day	Liver total P content					
		0-h	Mean	6-h	Mean	12-h	Mean
0IP/NP	10-d	1.02		0.91		1.04	
	20-d	1.06	1.08 <sup>a*</sup>	1.06	1.02 <sup>a</sup>	1.17	1.12 <sup>a</sup>
	30-d	1.18		1.08		1.14	
0IP/P	10-d	1.04		1.09		1.10	
	20-d	1.03	1.09 <sup>a</sup>	1.18	1.13 <sup>ab</sup>	1.19	1.15 <sup>ab</sup>
	30-d	1.21		1.12		1.17	
0.25IP/NP	10-d	1.09		1.08		1.12	
	20-d	1.17	1.15 <sup>ab</sup>	1.09	1.12 <sup>ab</sup>	1.15	1.16 <sup>ab</sup>
	30-d	1.19		1.21		1.20	
0.25IP/P	10-d	1.06		1.06		1.07	
	20-d	1.24	1.06 <sup>ab</sup>	1.29	1.16 <sup>b</sup>	1.21	1.17 <sup>ab</sup>
	30-d	1.17		1.13		1.22	
0.5IP/NP	10-d	1.13		1.04		1.18	
	20-d	1.27	1.13 <sup>b</sup>	1.3	1.15 <sup>b</sup>	1.29	1.23 <sup>c</sup>
	30-d	1.28		1.11		1.23	
0.5IP/P	10-d	1.16		1.11		1.19	
	20-d	1.26	1.22 <sup>b</sup>	1.29	1.21 <sup>b</sup>	1.19	1.22 <sup>bc</sup>
	30-d	1.25		1.24		1.27	

**ANOVA of Randomized Block**

Source	Post-prandial ALP (IU/dL)		
	0-h	6-h	12-h
Treatments	S	S	S
Block (day of sampling)	S	S	S
10 days	a*	a	a
20 days	b	b	b
30 days	b	b	b

\* Values in the each column with same superscripts are not significantly different (P>0.05)  
 S : Significant P< 0.05

absorption from plant ingredients which increased as fish weight increased to 10 g and then did not increase further in larger fish.

Different from the trend of postprandial plasma IP level, effect of phytase in increasing plasma Ca level was found only when dietary IP was not supplemented in diet. On the other hand, at higher dietary IP level, phytase supplementation did not significantly elevate plasma

Ca levels in all post-feeding time. In case of plasma Mg level, similar to plasma IP level, there was a clear trend of change of plasma Mg level at 1 and 3-h postprandial period due to the phytase supplementation. It is likely that Mg seemed to be more associated with the IP6 complex than Ca in Japanese flounder. This is in accordance with the previous studies on the dose-response relationship between di-

etary IP6 and mineral utilization using similar species that inclusion of dietary IP6 up to 13.5 g/kg diet did not significantly decrease the plasma Ca level (Laining *et al.*, 2010d). Similar finding was reported on Atlantic salmon that increasing of IP6 inclusion resulted in a progressive decrease of digestibility of Mg and Zn (Denstadli *et al.*, 2006). In contrast, Papatryphon *et al.* (1999) found that Ca was more associated with the phytate complex than other minerals in striped bass.

Post-prandial plasma ALP at 6-h period was strongly related to plasma IP level at the same post-feeding time. Regardless the dietary phytase, inclusion of dietary IP at 0.25% had a similar plasma ALP level to those fed diet without IP except at 3-h post-feeding time. However, increase of dietary IP to 0.5% significantly elevated plasma ALP level in all post-feeding time. In addition, plasma ALP significantly elevated by sampling days in all post-feeding time except at pre-feeding time. It is not known yet whether a relationship existed between the plasma ALP level and the activity of the digestive enzymes.

Even though ALP plays role in the absorption of several minerals in particular P and Ca for bone mineralization (Eguchi, 1995), effect of dietary IP and phytase supplement on plasma ALP in fish is so far still contradictory. Shearer & Hardy (1987) reported plasma ALP activity in rainbow trout was not significantly affected by feeding phosphorus sufficient and deficient diet. Since the importance of plasma ALP in fish nutrition, it is necessary to assess their variability with respect to physiological or environmental factors and determine the normal ranges of variation under different levels of such factors.

According to Lall (2002), absorbed P accumulates in skeletal tissue and soft tissue including liver, kidney, heart, muscle and blood. At pre and post-prandial liver P content, dietary 0.5% IP at two levels of phytase supplement significantly increased liver P content particularly at 12-h post-feeding time. Similar to plasma ALP level, accumulation of P in liver P seemed to be influenced by the sampling days which reflected the age of fish.

Change of 12-h postprandial Ca, Mg and ALP in other species after feeding dietary IP and phytase is extremely scarce. These preliminary results of blood sampling are useful

as additional information corresponding to varying effects depending on the dietary IP and phytase levels fed to Japanese flounder.

## CONCLUSION

Post-prandial plasma IP, Ca, Mg and ALP levels after 12-h feeding was lower in fish fed diet contained no dietary IP and phytase supplement. Supplementation of phytase at 0.25 and 0.5% IP had similar plasma IP, Mg and ALP levels at 6-h post-feeding. Maximum level of plasma IP and ALP appeared at 6-h post-feeding in fish fed diet supplemented with 0.25% IP together with phytase. Peak of plasma Mg level occurred at 3-h post-feeding in fish fed diet contained 0.25% IP together with phytase.

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