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## RECONFIRMING THE SPECIES OF MUD CRAB GENUS *SCYLLA* (DE HAAN, 1833) IN BALIKPAPAN, EAST KALIMANTAN PROVINCE, INDONESIA BASED ON MITOCHONDRIAL 16S rRNA

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(Received 24 April 2019; Final revised 23 April 2020; Accepted 23 April 2020)

### ABSTRACT

Taxonomy of mud crab species under the genus *Scylla* has been misidentified for several years due to their high morphological similarity. In Indonesia, some reports concerning mud crab have been published with misleading identification results where the species under the genus *Scylla* all named as *Scylla serrata*. The study was conducted to reconfirm the validity of species in the mud crab genus *Scylla* collected from Balikpapan mangrove, East Kalimantan, Indonesia and to analyze the genetic variation of the first generation (G-1) offspring, based on mitochondrial 16S rRNA sequence. The animal test used for species identification was a representative sample of mud crab. Ten of the G-1 crablet were randomly sampled for genetic variation analysis. Fragment of the 16S rRNA gene was isolated by PCR technique and purified for sequencing purpose. The mtDNA sequences were analyzed using Genetyx, BLAST-N, and DnaSP to get a consensus sequence, similarity index, haplotype and sequence diversity, and the number of haplotypes. The results showed that the 16S rRNA gene was successfully isolated with a single band in size of approximately 600 bp. The mud crab morphologically identified as *Scylla tranquebarica* was genetically confirmed as a species of *S. tranquebarica*. High haplotype diversity (0.9254) and low nucleotide diversity (0.1256) were revealed in the G-1 mud crab population, while the number of haplotypes was 7.5.

**KEYWORDS:** 16S rRNA; genetic variation; mitochondrial DNA; mud crab; species reconfirmation

### INTRODUCTION

Mud crab belonging to the genus *Scylla* is a commercially valuable crab resource for fisheries and aquaculture, both for local and international markets. Before the 1990s, most of the taxonomical works on the genus *Scylla* referred only to species of *S. serrata* (Mandal *et al.*, 2014a; Sarower *et al.*, 2016). The classification of genus *Scylla* has been controversial for a long time. Estampador (1949) claimed that based on the morphological characteristics, *Scylla* consisted of three species and one subspecies (four taxa) including *S. serrata*, *S. oceanica*, *S. tranquebarica*, and *S. serrata var. paramamosain*. However, the gene replacement at the three loci and the genetic distances between species suggested that the genus *Scylla* included at least three species (Fuseya & Watanabe, 1996).

Furthermore, Keenan *et al.* (1998) extensively revised the taxonomy of genus *Scylla* using integrative approaches (allozyme electrophoresis with sequencing cytochrome oxidase-I and 16s RNA). They divided this genus into four distinct species, including *S. serrata*, *S. olivacea*, *S. tranquebarica*, and *S. paramamosain*. The taxonomic status of genus *Scylla* based on morphological and molecular approaches, as proposed by Keenan *et al.* (1998), is widely accepted. The latest publication from Fazhan *et al.* (2020) hypothesized that the additional taxa in the genus *Scylla* with intermediate characteristics could be presumed hybrids. However, this work needs to be validated at the molecular level. Up to the present, misidentification of the species in genus *Scylla* is still a big problem related to developmental stage, size difference, and close similarity.

Species identification is a starting point in bio-ecological studies, and correctly-determined species is fundamental to develop a breeding program. The high morphological diversity and plasticity of mud crabs due to developmental stages and environment

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have made it difficult to accurately identify a species of mud crabs based only on morphological characteristics (Balasubramanian *et al.*, 2014; Mandal *et al.*, 2014a, 2014b; Sarower *et al.*, 2016).

DNA-based identification techniques have been developed and proven to be analytically robust. A short DNA sequence of the mitochondrial genome area could be taken as a DNA barcode for species identification. A gene of 16S rRNA is one of the most common mitochondrial DNA (mtDNA) gene markers used to identify species because it belongs to the most conservative gene among the mtDNA protein-coding genes (Yang *et al.*, 2014). Eventhough mitochondrial 16S ribosomal RNA gene regions have been used in genetic studies for various purposes, (Imai *et al.*, 2004; Masaoka & Kobayashi, 2005; Akasaki *et al.*, 2006; Saavedra & Pena, 2004; Srinu *et al.*, 2018), their usage in species identification at a molecular level for mud crab is still very limited. The study aimed to reconfirm the validity of species in the mud crab genus *Scylla* collected from Balikpapan mangrove, East Kalimantan, Indonesia and to analyze the genetic variation of the first generation (G-1) offspring, based on mitochondrial 16S rRNA sequence.

## MATERIALS AND METHODS

### Sample Collection

Mud crabs ( $n=30$ ) with a weight of  $235.7 \pm 88.4$  g were collected from a mangrove area surrounding brackish water ponds along the coastline of Balikpapan Bay, Balikpapan, East Kalimantan. The mud crabs were morphologically identified as a species of *S. tranquebarica* (Gunarto *et al.*, 2017). One individual sample weighed 228.3 g was randomly taken to re-identify its species using the molecular technique. The other mud crabs were continuously reared to produce the first generation (G-1) offsprings via individual spawning. The gonadal maturation of broodstock was performed based on the standard operating procedure (SOP) established in mud crab hatchery of Research Institute for Brackishwater Aquaculture and Fisheries Extension (RIBAFE) in Barru, South Sulawesi (see Gunarto *et al.*, 2016). After hatching, the larvae from each parent were reared separately in a conical fiber tank until reaching the crablet stage and then harvested at crablet D-10. Furthermore, the crablets were reared in a concrete tank until reaching crablet D-30 with a weight range of 1-2 g. Ten individual crablets were randomly collected from six parents for genetic analysis purposes.

### Genomic DNA Extraction

The genomic DNA of adult and mud crab crablet (G-1) was isolated using the phenol-chloroform extraction method developed by Parenrengi *et al.* (2000).

The purity of genomic DNA was estimated by measuring in a GeneQuant. The DNA concentration and purity were quantitatively determined from the ratio between the reading of absorbance at 260 nm and 280 nm ( $OD_{260}/OD_{280}$ ) (Linacero *et al.*, 1998). DNA purity was qualitatively observed through the appearance of the band on the agarose gel.

### Isolation and Sequencing of 16S rRNA Gene

A gene encoding mtDNA 16S rRNA of mud crab was isolated according to the method of Tenriulo *et al.* (2009). The method has been applied to green tiger prawn *Penaeus semisulcatus* by using a pair of specific primers, i.e., 16SrRNA-F: 5'- cgc ctg ttt aac aaa aac at -3' and 16SrRNA-R: 5'- ccg gtc tga act cag atc atg t -3'. PCR reaction consisted of PureTaq Ready-To-Go PCR Beads kit, 1  $\mu$ L for each primer (50  $\mu$ mol), and sterile water until the final volume of 25  $\mu$ L. PCR was processed in the 2720 Thermal Cycler with the following program: initial denaturation of 15 min at 55°C and 2 min at 94°C; followed by 40 cycles of denaturation 15 sec at 94°C, annealing 30 sec at 60°C, and extension 1 min 15 sec at 68°C; and final extension of 5 min at 72°C. The success of mtDNA isolation was determined by running the PCR product at 0.8% agarose gel to visualize the single fragment result. The good PCR product was purified using commercial Kit and DNA concentration was measured by using GeneQuant before the PCR products were sent to the First Base for reading nucleotide sequences by ABI sequence scanner.

### Data Analysis

The mtDNA sequence results were analyzed by using Genetyx Version 7.0, BLAST-N (basic local alignment search tool-nucleotide) (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>) and DnaSP Version 5.0 (Librado & Rozas, 2009). The Genetyx program was applied to obtain the consensus of forward-reverse sequences and determine nucleotide variation of intra-population. BLAST-N was used to confirm species identification by sequence analysis of similarity index among crustacean species deposited at GenBank. DnaSP was performed to get the haplotype variation and number of mud crab larvae.

## RESULTS AND DISCUSSION

### Genomic DNA and 16S rRNA Isolation

Genomic DNA of mud crab was successfully extracted using the conventional method in a concentration of  $34.5 \pm 11.4$   $\mu$ g mL<sup>-1</sup> with a high purity level of  $1,873 \pm 0,099$ . Linacero *et al.* (1998) recommended the DNA purity for molecular analysis in the range of 1.8-2.0. This method could also produce a high purity of genomic DNA on tiger shrimp (Tenriulo *et al.*,

2012; Parenrengi *et al.*, 2017) and rabbitfish (Lante *et al.*, 2011). A partial 16S rRNA gene was successfully isolated from the genomic DNA of mud crab with a single band in size of approximately 600 bp (in range of 500-750 bp) (Figure 1). This range of DNA fragment position of the mtDNA gene was also observed in green tiger prawn *P. semisulcatus* (Tenriulo *et al.*, 2009).

**Confirmation of Mud Crab Species Identification**

A consensus sequence in the length of 568 bp (Figure 2) was used as a reference to confirm the species identification of mud crab understudy at the molecular level by BLAST-N online. At least 116 mtDNA sequences of crustacean species at GenBank exhibited a high similarity in the range of 93-100% with

the species in the present study. Twenty representatives of mtDNA genes of mud crab with high similarity were used as references and listed in Table 1. The mud crab sample had an identical sequence of mtDNA genes (100% similarity) to *S. tranquebarica* isolate of P-ST-01, a partial sequence of 16S rRNA gene (accession number of KT921342.1). So, it is argued here that the mud crab sample in this study was unmistakably confirmed as a species of *S. tranquebarica*. This species identification was accurately supported by the high similarity (identical sequence) and the rate of maximum scoring (total score). The sequence similarity of the 16S rRNA gene at GenBank was very close to species of *S. tranquebarica*, followed by *S. olivacea* (99%), *S. paramamosain* (95%), and *S. serrata* (93%). The highest scoring value was obtained by species of *S. tranquebarica* (1,022);

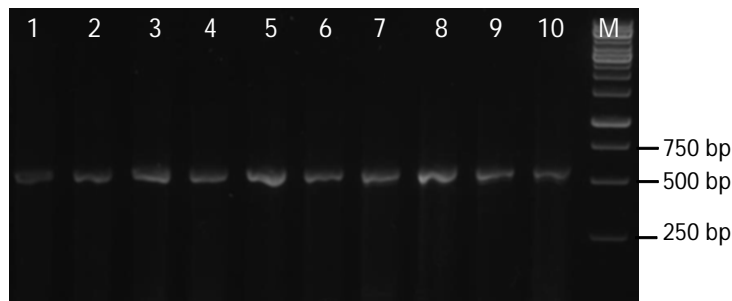


Figure 1. The single band of mitochondrial 16S rRNA gene isolated from the G-1 offspring of mud crab on 0.8% agarose gel (M=DNA marker and 1-10=crablet samples).

10	20	30	40	50	60
TTAAATCGCC	TGTTTTACAA	AAACATGTCT	ATTTGAAATT	ATATATAGTC	TAGCCTGCTC
70	80	90	100	110	120
ACTGACAAAG	ATGTTAAATA	GCCGCGGTAT	TTTGACCGTG	CAAAGGTAGC	ATAATCATT A
130	140	150	160	170	180
GTTTTTTAAT	TGGAACCTTG	TATGAATGGT	TGGACAAAGA	AAAAGCTGTC	TCTTTTATTA
190	200	210	220	230	240
AAATTGAAAT	TAACCTCTAA	GTGAAAAGGC	TTAGATTTTC	TAGAGGGACG	ATAAGACCCT
250	260	270	280	290	300
ATAAAGCTTT	ATAAATTAAG	TAAGTTAGAA	CAAATTTTAT	AGAATAAAGG	TAAATTTAAT
310	320	330	340	350	360
ATATTTATTT	GGTTGGGGCG	ACAATGGTAT	AAATTAAATA	ACTGCTATTA	AAATTAACA
370	380	390	400	410	420
ATTATATTTG	ATTAAAATTA	TAATTGATCC	TTATTATGGA	TTAAAAGATC	AAGTTACTTT
430	440	450	460	470	480
AGGGATAACA	GCGTAATTTT	TTTTAAGAGT	TCTTATCAAA	GAAGAAGTTT	GCGACCTCGA
490	500	510	520	530	540
TGTTGAATTA	AAATGTCTTT	ATAGTGCAGA	AGCTATAAAA	GAAGGTCTGT	TCGACCTTTA
550	560	570			
ATTTTTTACA	TGATCTGAGT	TCAGACCG			

Figure 2. Partial nucleotide sequences of gene encoding the mitochondrial 16S rRNA isolated from mud crab.

Table 1. Sequence similarity and total score of mitochondrial 16S rRNA gene isolated from mud crab

Description	Total score	Similarity (%)	Accession
<i>Scylla tranquebarica</i> isolate P-ST-01 16S ribosomal RNA gene, partial sequence; mitochondrial	963	100	KT921342.1
<i>Scylla tranquebarica</i> 16S ribosomal RNA gene, partial sequence	1.022	99	AF109320.1
<i>Scylla tranquebarica</i> mitochondrion, complete genome	1.02	99	FJ827759.1
<i>Scylla tranquebarica</i> voucher DLSU.MI.00032 16S ribosomal RNA gene, partial sequence; mitochondrial	953	99	KM258653.1
<i>Scylla tranquebarica</i> voucher DLSU.MI.00033 16S ribosomal RNA gene, partial sequence; mitochondrial	950	99	KM258654.1
<i>Scylla olivacea</i> isolate SOPB4 16S ribosomal RNA gene, partial sequence; mitochondrial	935	99	KU306796.1
<i>Scylla olivacea</i> isolate SOLG5 16S ribosomal RNA gene, partial sequence; mitochondrial	935	99	KU306746.1
<i>Scylla olivacea</i> mitochondrion, complete genome	824	93	FJ827760.1
<i>Scylla olivacea</i> 16S ribosomal RNA gene, mitochondrial gene for mitochondrial RNA, partial sequence	821	93	AF109321.1
<i>Scylla olivacea</i> voucher NBFGR-CHN-Scylla1 16S ribosomal RNA gene, partial sequence; mitochondrial	813	93	KC154070.1
<i>Scylla paramamosain</i> 16S ribosomal RNA gene, partial sequence	893	95	AF109319.1
<i>Scylla paramamosain</i> mitochondrion, complete genome	891	95	MG197997.1
<i>Scylla paramamosain</i> mitochondrion, complete genome	891	95	JX457150.1
<i>Scylla paramamosain</i> mRNA sequence	891	95	HM217900.1
<i>Scylla paramamosain</i> mitochondrion, complete genome	891	95	FJ827761.1
<i>Scylla serrata</i> mitochondrion, complete genome	824	93	HM590866.1
<i>Scylla serrata</i> 16S ribosomal RNA gene, mitochondrial gene for mitochondrial RNA, partial sequence	815	93	AF109318.1
<i>Scylla serrata</i> mitochondrion, complete genome	813	93	FJ827758.1
<i>Scylla serrata</i> voucher NBFGR-CHN-Scylla10 16S ribosomal RNA gene, partial sequence; mitochondrial	811	93	KC154088.1
<i>Scylla serrata</i> voucher NBFGR-CHN-Scylla8 16S ribosomal RNA gene, partial sequence; mitochondrial	808	93	KC154086.1

followed by *S. olivacea* (935); *S. paramamosain* (893); and *S. serrata* (824).

Prior to this study, Imai *et al.* (2004) had reported species identification for four species of mud crabs using RFLP (*restriction fragment length polymorphism*) mtDNA marker, especially for enzyme restriction of *Dra-I* and *Hind-III*. Species confirmation of mud crab conducted by Sarower *et al.* (2016) using the mtDNA marker showed that mud crabs collected in Bangladesh were identified as a species of *S. olivacea*. Their result challenged erroneous species identification results from other studies that mistakenly identified the species as *S. serrata*. The mud crab *S. serrata* commonly mentioned in India's literature was, in fact, *S. olivacea* when identified using RAPD and PCR-RFLP markers (Mandal *et al.* 2014a) and sequencing of COI gene (Mandal *et al.*, 2014b). In addition, a comprehensive DNA sequence analysis of mud crabs in India

revealed that the species commonly reported as *S. tranquebarica* was, in fact, *S. serrata* (Balasubramanian *et al.*, 2014). Based on the evidence from this present study, we argue that the mud crab of *S. serrata* commonly reported in Indonesia's past studies is likely to be a species of either *S. olivacea* or *S. tranquebarica*.

#### Genetic Variation of The G-1 Mud Crab *Scylla tranquebarica*

The reading of mtDNA 16S rRNA nucleotide sequences of mud crabs was in the range of 573-624 bp. The sequence alignment of ten mud crab larvae showed that the reading of forward and reverse sequences was not started in the same position, while the variation of nucleotide was in sequence position of 26, 27, 30, 39, 516, 558, 574, and 595 bp (Figure 3).

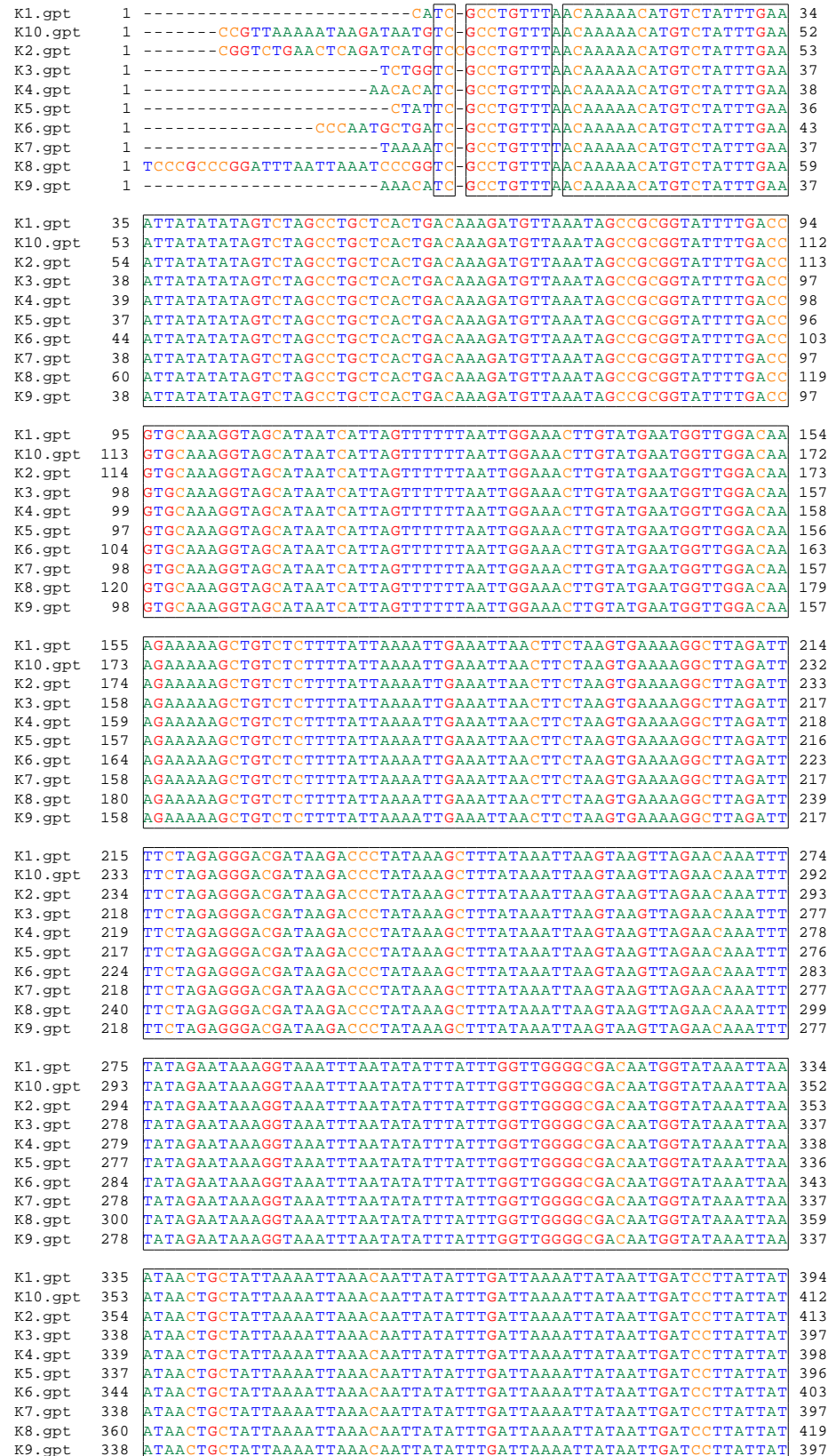


Figure 3. Alignment of mtDNA 16S rRNA sequence of mud crab on 1<sup>st</sup> generation (F1) from Balikpapan broodstocks (A= adenine; T= thymine, G= guanine, C= cytosine, and without box indicating the nucleotide variation).

K1. gpt	395	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	454
K10. gpt	413	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	472
K2. gpt	414	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	473
K3. gpt	398	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	457
K4. gpt	399	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	458
K5. gpt	397	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	456
K6. gpt	404	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	463
K7. gpt	398	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	457
K8. gpt	420	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	479
K9. gpt	398	GGATTA AAAAGATCAAGTTACTTTAGGGATAACAGCGTAATTTCTTTTAAGAGTTCTTATC	457
K1. gpt	455	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	514
K10. gpt	473	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	531
K2. gpt	474	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	532
K3. gpt	458	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	516
K4. gpt	459	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	517
K5. gpt	457	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	515
K6. gpt	464	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	522
K7. gpt	458	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	516
K8. gpt	480	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	538
K9. gpt	458	AAAGAA GAAGTTTGGACCTCGATGTTGAATTAATAATGTCCTTTATAGTGCAGAAGCTAT	516
K1. gpt	515	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGAAAAGTC	573
K10. gpt	532	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGACATGC	590
K2. gpt	533	AAAA GAAGGTCTGTTCCGACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGG----	587
K3. gpt	517	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGACTT--	573
K4. gpt	518	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGATATAT	576
K5. gpt	516	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGATTTCT	574
K6. gpt	523	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGA--CTC	579
K7. gpt	517	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGATTTTT	575
K8. gpt	539	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGAGACAA	597
K9. gpt	517	AAAA GAAGGTCTGTTCC-ACCTTTAATTTTTTACATGATCTGAGTTTCAGACCGGATTTTA	575
K1. gpt	574	CATG-----	577
K10. gpt	590	-----	590
K2. gpt	587	-----	587
K3. gpt	573	-----	573
K4. gpt	577	CATGATCTGAGTTC-----	590
K5. gpt	575	CATG-----	578
K6. gpt	580	CATGATC-----	586
K7. gpt	576	CT-----	577
K8. gpt	598	CATGATCTGAGTTCAGACCGGACTTGA	624
K9. gpt	576	CT-----	577

Figure 3. Continued

The genetic variation analysis of the mud crab larvae population revealed haplotype variation as much as 0.9254, and the number of haplotypes was 7.5, with a nucleotide variation of 0.1256. The high haplotype diversity observed in this present study indicated the high genetic variation of mud crab *S. tranquebarica* population in Balikpapan, in which these data are useful information for mud crab breeding program in the future. In contrast, Sharif *et al.* (2016) reported the lower haplotype variation of the mtDNA CO-I gene in 5 populations of mud crab *S. tranquebarica* in Sabah, Malaysia, where the averages of haplotype variation and nucleotide were of 0.5564 and 0.0038, respectively. Different approaches using microsatellite markers on mud crab, *S. paramamosain* in China, had reported high heterozygosity of 0.62-0.67 (Ma *et al.*, 2012).

Other studies also reported similar findings regarding the high haplotype and low nucleotide variations in mud crab *S. olivacea*. Rosly *et al.* (2013) reported that high haplotype diversity (0.968) and low nucleotide diversity (0.120) were displayed in mud

crab populations of the east and west coast of Malaysia Peninsular. A haplotype is one of the biological indicators which can be used to analyze the genetic variation of a population. The high haplotype variation of mud crab in the present study offers a significant advantage to develop a breeding program of the crab species in the future. Imai *et al.* (2002) have successfully used D-loop mtDNA haplotype as an indicator to measure the stocking affectivity of wild populations of mud crab *S. paramamosain*. The mtDNA marker could act as a DNA barcode for animal species identification due to more straightforward isolation procedures and good resolution results for animal species identification. The other markers, such as RAPD, have been used to assess the genetic variation of three mud crab species, *S. serrata*, *S. oceanica*, and *S. tranquebarica* mud crab. Klinbunga *et al.* (2000) reported that the percentage of polymorphic bands within these populations ranged from 47.92% to 77.59%, and species-specific RAPD markers were also observed and used to construct a molecular diagnostic key in these taxa.

Knowledge of genetic diversity is an essential aspect of ensuring sustainable exploitation of any commercial fish species. The findings from the present study are arguably important not only in providing baseline information on taxonomic identification and genetic diversity of *S. tranquebarica* species but also in the overall management of mud crabs for fisheries and aquaculture uses considering the increasing market demand for this species.

## CONCLUSION

This study found that the mud crab understudy was a species of *S. tranquebarica* confirmed through its 16S rRNA gene, which was identical to the mtDNA sequence (100%) of the same species at GenBank. The haplotype variation of 0.9254, sequence nucleotide variation of 0.1256, and haplotype number of 7.5 were observed in the populations of G-1 mud crab *S. tranquebarica*. The high haplotype diversity of the mud crabs suggested a potential quantitative trait for the breeding program of this species. However, the proper species identification should be performed before developing the mud crab culture in the future.

## ACKNOWLEDGMENT

We gratefully acknowledge the support of the Ministry of Marine Affairs and Fisheries, the Republic of Indonesia, for funding this study through the 2017 DIPA-RIBAFE. We also thank the researchers and technicians at the Biotechnology Laboratory in RIBAFE, Maros, and the technicians of crab hatchery in Barru for their research assistance.

## REFERENCES

- Akasaki, T., Saruwatari, T., Tomonaga, H., Sato, S., & Watanabe, Y. (2006). Identification of imported Chirimén at the genus level by a direct sequencing method using mitochondrial partial 16S rDNA region. *Fisheries Science*, 72, 686-682.
- Balasubramanian, C.P., Cubelio, S.S., Mohanlal, D.L., Ponniah, A.G., Kumar, R., Bineesh, K.K., Ravichandran, P., ....., & Jena, J.K. (2014). DNA sequence information resolves taxonomic ambiguity of the common mud crab species (Genus *Scylla*) in Indian waters. *Mitochondrial DNA*, DOI: 10.3109/19401736.2014.892076.
- Estampador, E.P. (1949). Studies on *Scylla* (Crustacea:Portunidae) I. Revision of the genus. *The Philippines Journal of Science*, 78, 95-109.
- Fazhan, H., Waiho, K., Qunitio, E., Baylon, J.C., Fujaya, Y., Rukminasari, N., Azri, M.F.D., ....., & Ikhwanuddin, M. (2020). Morphological descriptions and morphometric discriminant function analysis reveal an additional four groups of *Scylla* spp. *PeerJ*, 8: e8066. DOI: 10.7717/peerj.8066.
- Fuseya, R. & Watanabe, S. (1996). Genetic Variability in the Mud Crab Genus *Scylla* (Brachyura: Portunidae). *Fisheries Science*, 62(5), 705-709.
- Gunarto, Syafaat, M.N., Herlinah, Parenrengi, A., & Mustafa, A. (2016). *Biological Aspect and seed production technique of mud crab Scylla spp* [in Indonesian]. Technical Book of Research Institute for Coastal Aquaculture, 62 pp.
- Gunarto, Sulaeman, Tjomba, H., Syafaat, M.N., & Nurbaya. (2017). *Improvement of production system and broodstock candidate for mud crab domestication* [in Indonesian]. Technical Report of Research Institute for Brackishwater Aquaculture and Fisheries Extension, 23 pp.
- Imai, H., Obata, Y., & Sekiya, S. (2002). Mitochondrial DNA markers confirm successful stocking of mud crab juveniles, (*Scylla paramamosain*) into a natural population. *Suisanzoshoku*, 50(2), 149-156.
- Imai, H., Cheng, J.H., Katsuyuki, Hamasaki, K., & Numachi, K.I. (2004). Identification of four mud crab species (genus *Scylla*) using ITS-1 and 16S rDNA markers. *Aquatic Living Resource*, 17, 31-34.
- Keenan, C.P., Peter, J.F.D., & Mann, D.L. (1998). A revision of the genus *Scylla* De Haan, 1833 (Crustacea: Decapoda: Brachyura: Portunidae). *The Raffles Bulletin of Zoology*, 46(1), 217-245.
- Klinbunga, S., Boonyapakdee, A., & Pratoomchat, B. (2000). Genetic diversity and species-diagnostic markers of mud crabs (Genus *Scylla*) in Eastern Thailand determined by RAPD analysis. *Marine Biotechnology*, 2(2), 180-187.
- Lante, S., Tenriulo, A., Parenrengi, A., Syah, R., & Malina, A.C. (2011). Determining population genetic variability of rabbitfish (*Siganus guttatus*) from Makassar Strait and Bone Bay waters using random amplified polymorphic DNA method. *Jurnal Riset Akuakulture*, 6(2), 211-224.
- Librado, P. & Rozas, J. (2009). DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. *Bioinformatic*, 25, 1451-1452.
- Linacero, R., Rueda, J., & Vazquez, A.M. (1998). Quantification of DNA. In *Molecular tools for screening biodiversity: plants and animals* (p.18-21). London, Weinheim, New York, Tokyo, Melbourne, Madras: Chapman and Hall.
- Ma, H., Cui, H., Ma, C., & Lingbo Ma, L. (2012). High genetic diversity and low differentiation in mud crab (*Scylla paramamosain*) along the south-eastern coast of China revealed by microsatellite

- markers. *The Journal of Experimental Biology*, 215, 3120-3125.
- Mandal, A., Varkey, M., Mani, A.K., Sobhanan, S.P., & Thampi-Samraj, Y.C. (2014a). Identification of Indian Mangrove Mud Crab Genus *Scylla* spp. (Brachyura: Portunidae) using RAPD and PCR-RFLP Markers. *Journal of Shellfish Research*, 33(2), 503-510.
- Mandal, A., Varkey, M., Sobhanan, S.P., Mani, A.K., Gopalakrishnan, A., Kumaran, G., Sethuramalingam, A., ....., & Samraj, Y.C. (2014b). Molecular markers reveal only two mud crab species of genus *Scylla* (Brachyura: Portunidae) in Indian coastal waters. *Biochemical Genetics*, 52(7-8), 338-354.
- Masaoka, T. & Kobayashi, T. (2005). Species identification of *Pinctada imbricate* using intergenic spacer of nuclear ribosomal RNA gene and mitochondrial 16S ribosomal RNA gene regions. *Fisheries Science*, 71, 837-846.
- Paran, F.J. (2016). Phylogeography and population genetic of the orange mud crab, *Scylla Olivacea*, in *Philippine waters*. University of the Philippines.
- Parenrengi, A., Shamsudin, L., Ismail, P., & Amin, N.M. (2000). Preliminary study on DNA level marker of grouper at different buffer preservation and DNA extraction method. In Saad MS, Faridah QZ, Kadir MA., Khalid MZZ, Mohamad O, Saleh GB, Panandam JM (Editors). *Genetic Manipulation: Challenges and Advantages*. Proceeding of the 4<sup>th</sup> National Congress on Genetics, 26-28 Sept.2000, Genting Highlands, Malaysia, p. 194-208.
- Parenrengi, A., Alimuddin, & Tenriulo, A. (2017). Characteristics of viral protein, VP-15, of whitespot syndrome virus isolated from infected tiger shrimp *Penaeus monodon* (Fabricius, 1798). *Indonesian Aquaculture Journal*, 12(2), 67-75.
- Rosly, H.A.A.M., Nor, S.A.M., Yahya, K., & Naim, D.M. (2013). Mitochondrial DNA diversity of mud crab *Scylla olivacea* (Portunidae) in Peninsular Malaysia: a preliminary assessment. *Molecular Biology Reports*, 40(11), 6407-6418.
- Saavedra, C. & Pena, J.B. (2004). Phylogenetic relationships of commercial European and Australasian king scallops (*Pecten* spp.) based on partial 16S ribosomal RNA gene sequences. *Aquaculture*, 235, 153-166.
- Sharif, N.A.M., Kahar, N.A.S., Rodrigues, K., Ransangan, J., & Kian, A.Y.S., (2016). Genetic diversity of mud crabs, *Scylla tranquebarica* in Sabah, Malaysia based on Cytochrome C Oxidase (COI) gene sequence. *Songklanakarin Journal of Science and Technology*, 8(4), 365-372.
- Sarower, M.G., Shahriar, S.I.M., Nakamura, H., Rouf, H.A., & Okada, S. (2016). Taxonomic confirmation of mud crab species (genus *Scylla*) in Bangladesh by nuclear and mitochondrial DNA markers. *Journal Mitochondrial DNA Part A, DNA Mapping, Sequencing, and Analysis*, 28(6), 935-940.
- Srinu, G., Rao, G.L., SK Chand Basha, S.C., & Pandayya, B. (2018). Molecular identification and validation of Indian mud crab genus *Scylla* (Decapoda: Portunidae) based on mitochondrial genes. *International Journal of Zoology Studies*, 3(1), 96-102.
- Tenriulo, A., Parenrengi, A., & Sulaeman. (2009). *Genetic variation of green tiger prawn Penaeus semisulcatus based on the mitochondrial DNA* [Indonesian]. In National Seminar of Fisheries, Fisheries High School.
- Tenriulo, A., Parenrengi, A., & Alimuddin. (2012). Analysis of microsatellite marker of diseases resistant tiger shrimp *Penaeus monodon* from several wild populations [in Indonesian]. In *Seminar Nasional Tahunan IX Hasil Penelitian Perikanan dan Kelautan Tahun 2012*, Fisheries Department, Agriculture Faculty, Gajah Mada University.
- Yang, L., Tan, Z., Wang, D., Xue, L., Guan, M.X., Huang, T., & Lia, R. (2014). Species identification through mitochondrial rRNA genetic analysis. *Scientific Reports*, 4: 4089, doi: 10.1038/srep04089.